



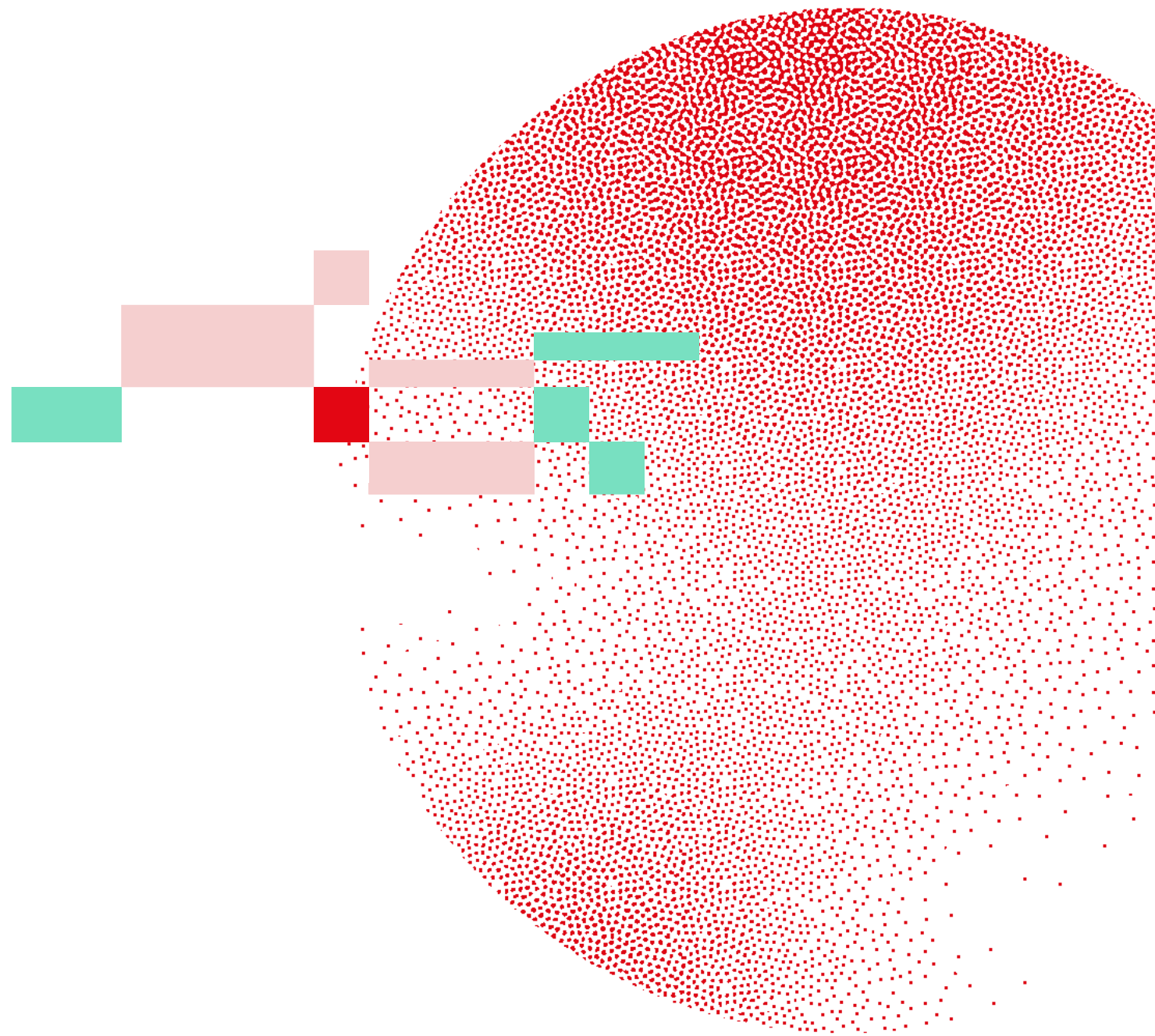
Swiss Institute of  
Bioinformatics

DAY 2, PART 2

# AlphaFold3

Diana Rapota, Rok Breznikar, Janani Durairaj

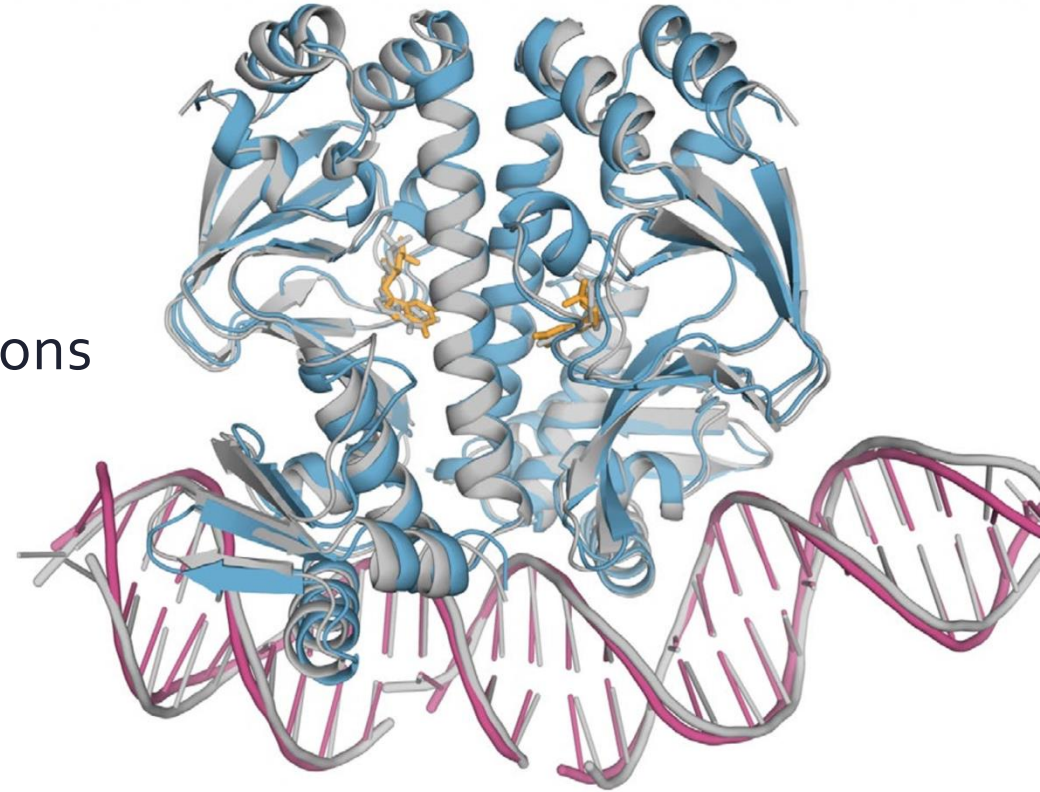
23-24 June 2026





# What more you can do with it

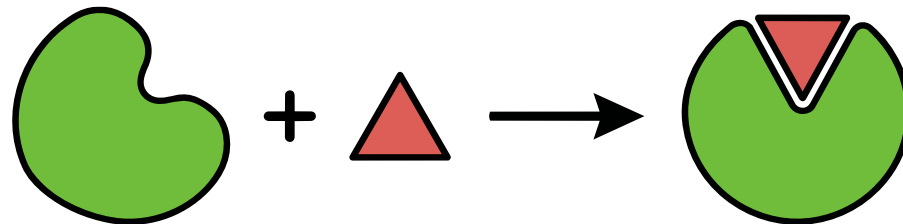
- Proteins
- DNA, RNA
- Small molecules
- Ions
- Modified residues, post-translational modifications
- Complexes of all of these
- Doesn't model:
  - Water
  - Hydrogens





# Protein-ligand complex

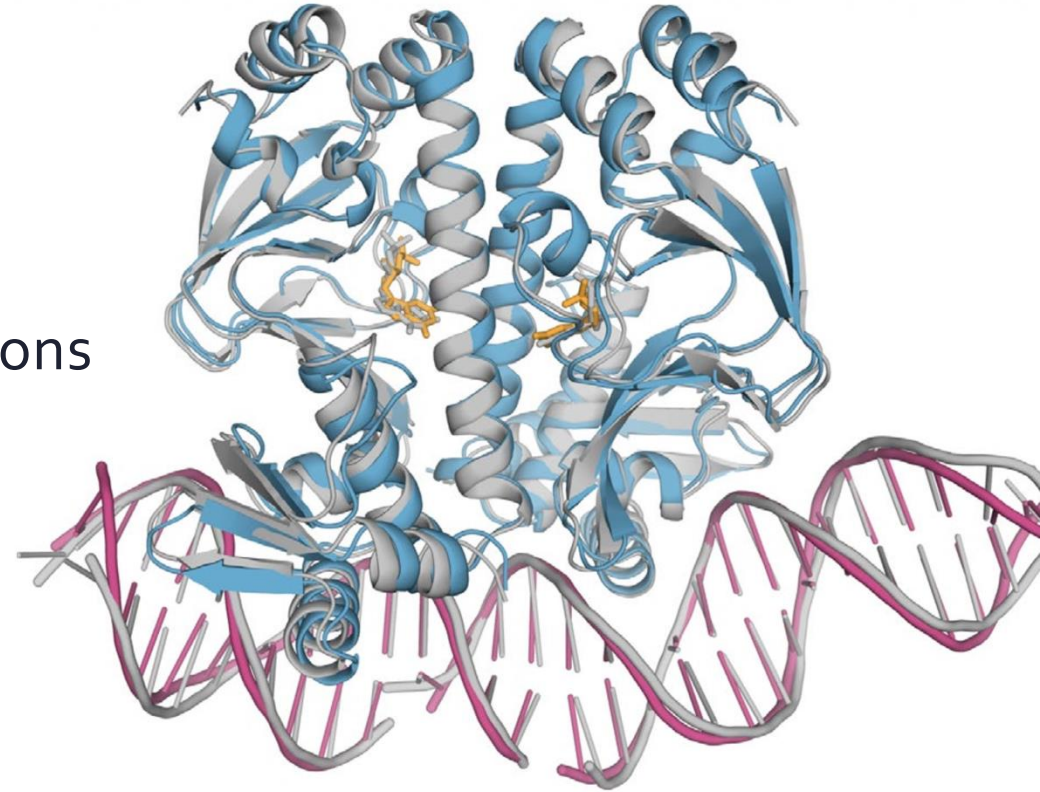
- Taking AlphaFold2 structure (protein on its own) and adding the ligand doesn't work that well (docking)
- Induced fit  
(protein is not in the correct conformation for the ligand)





# What more you can do with it

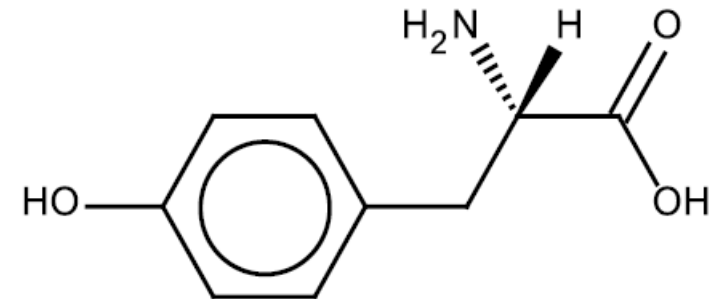
- Proteins
- DNA, RNA
- Small molecules
- Ions
- Modified residues, post-translational modifications
- Aims to model the whole complex together





# SMILES

- String notation for small molecules (ligands)
- Describes a molecular graph with 4 basic rules
  - Atoms
  - Bonds
  - Branching
  - Ring closures
- Hydrogens can be included or omitted (typically omitted)





# SMILES

## Atoms

C

O

N

...

## Bonds

- (single)  
(usually omitted)

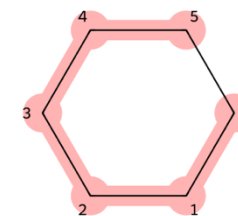
= (double)

# (triple)

## Branching

( )

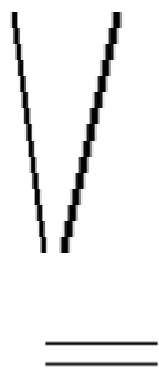
## Ring closure numbers



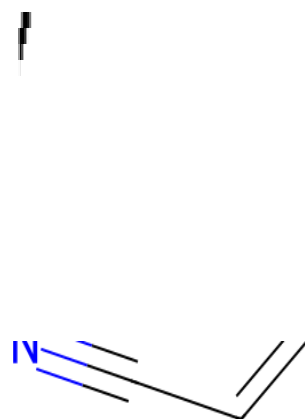
C1CCCCC1



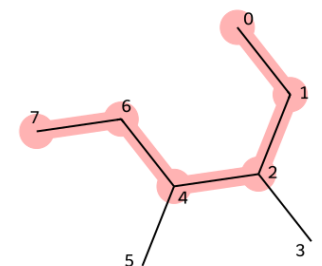
C=C



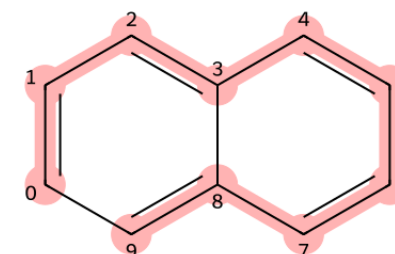
C#C



C=CC#N



CCC(C)C(C)CC



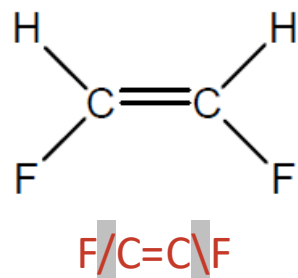
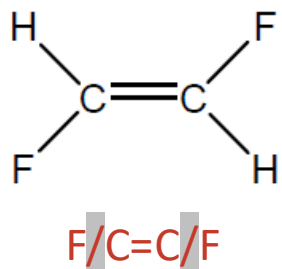
c1ccc2ccccc2c1



# SMILES

## Orientation around double bonds

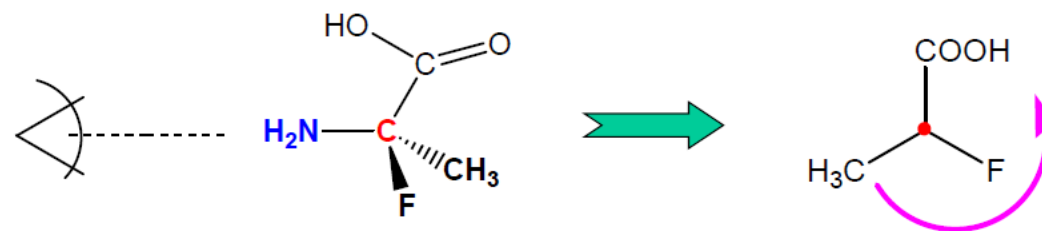
/ and \



## Chiral centers

@: ↻

@@: ↺



N[C@](C)(F)C(=O)O



# Tokenization

AlphaFold2 only dealt with proteins

→ divided into residues



# Tokenization

AlphaFold3 deals with more

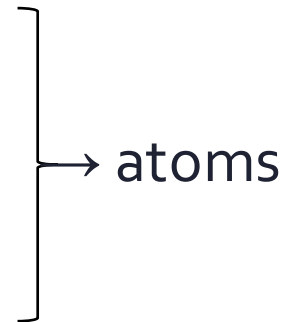
Proteins → residues

DNA, RNA → nucleotides

Small molecules

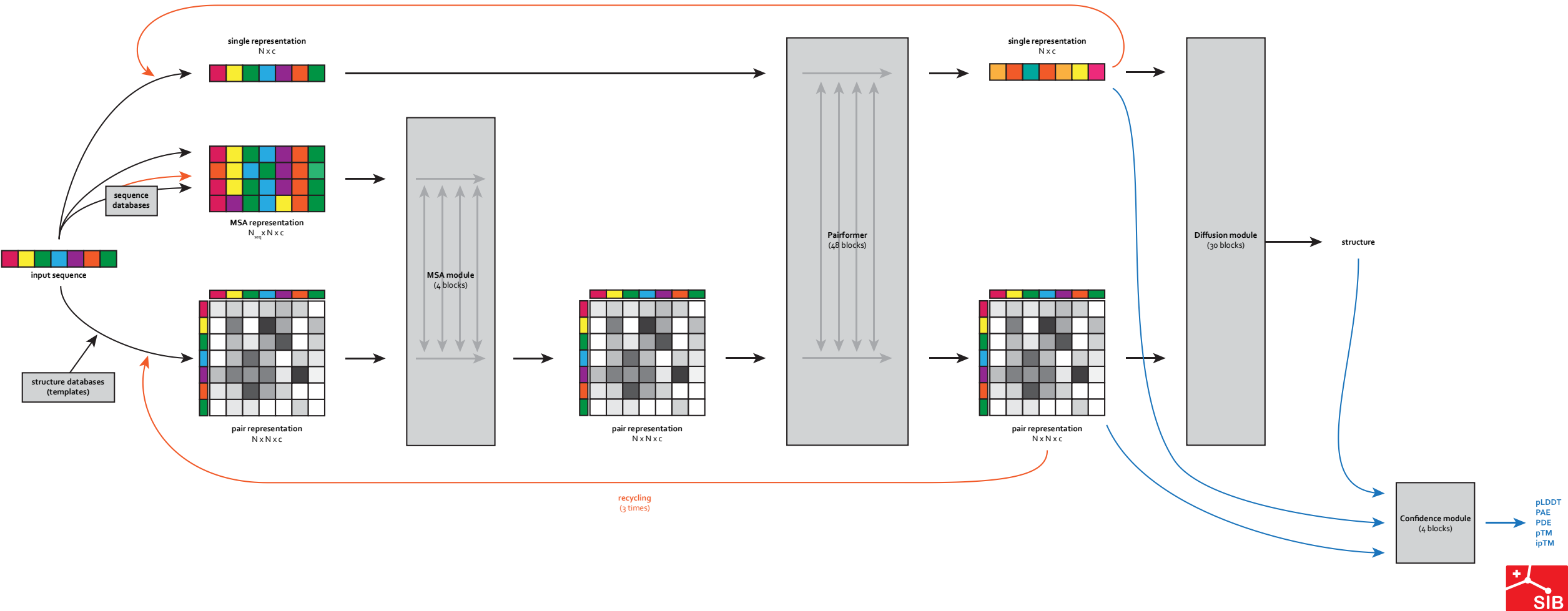
Ions

Modified residues

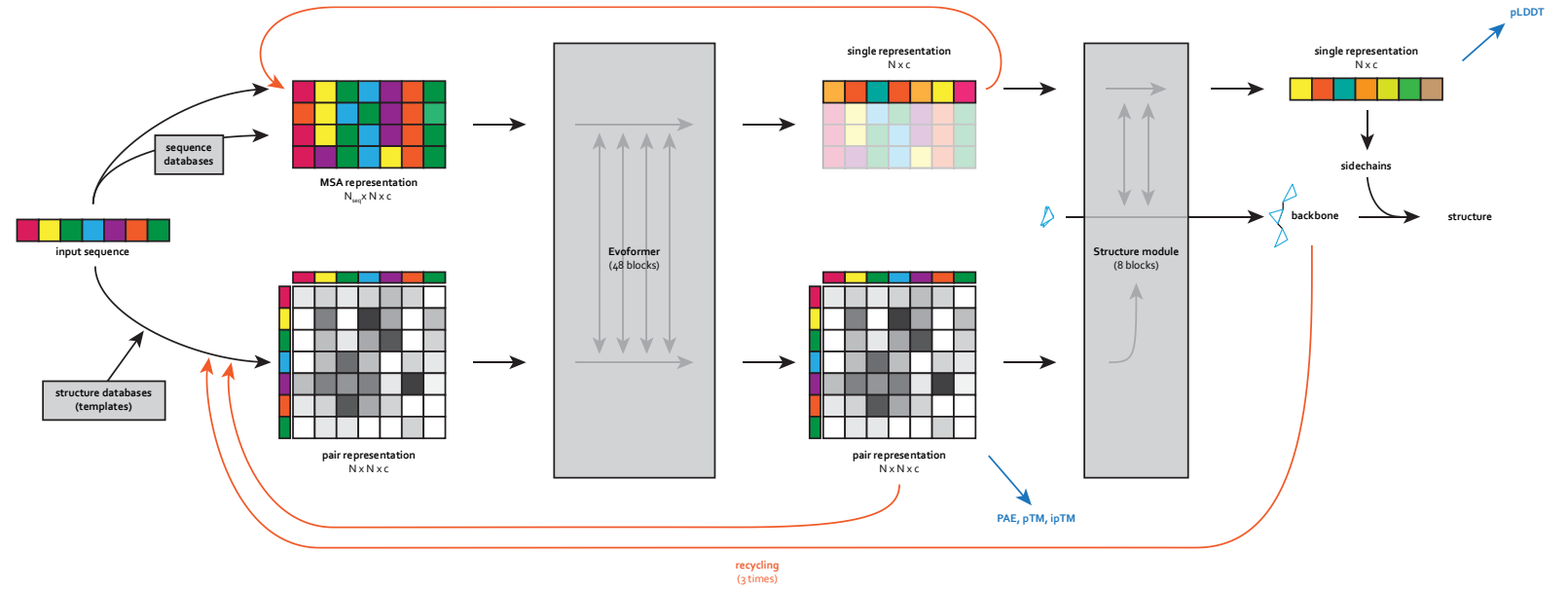


# Architecture

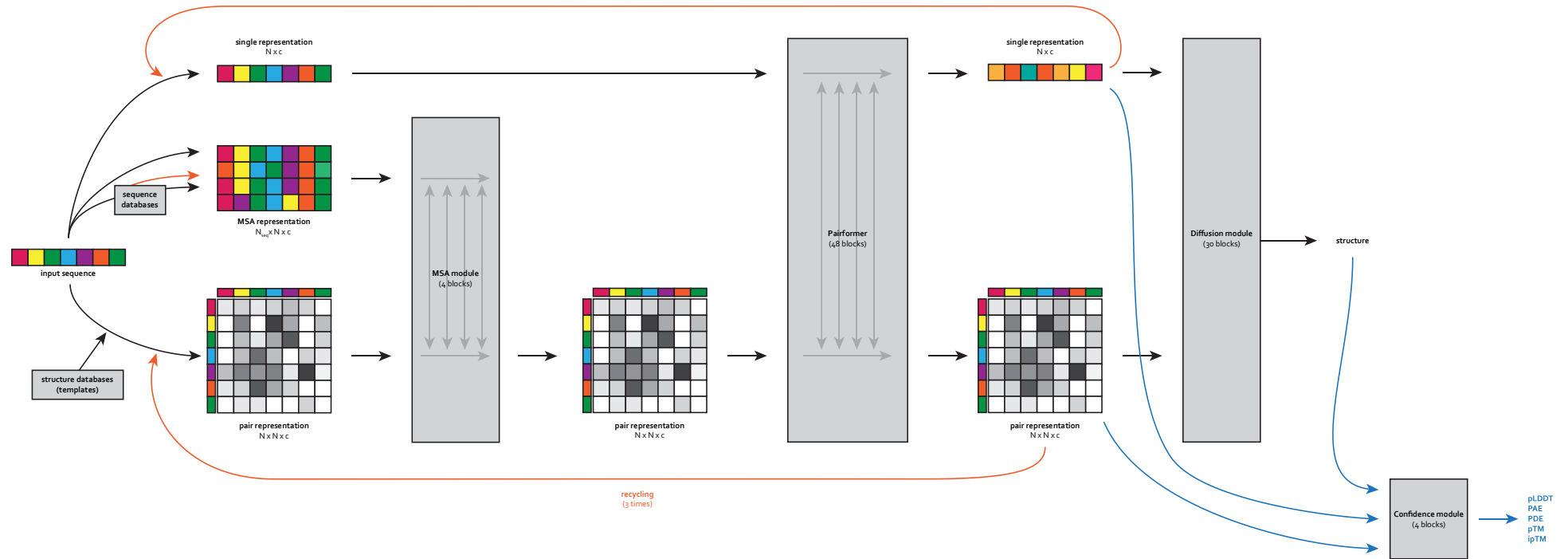
- Simplified scheme (conversions between atom and token level representations)



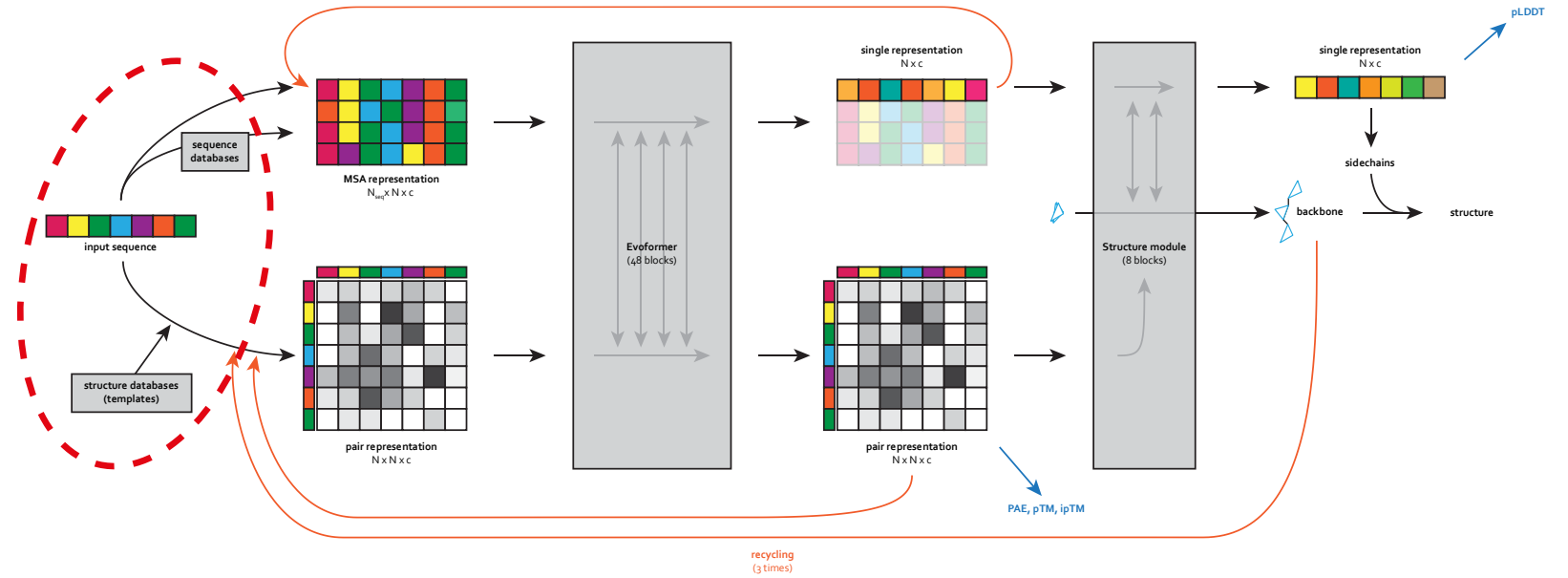
# AlphaFold2



# AlphaFold3

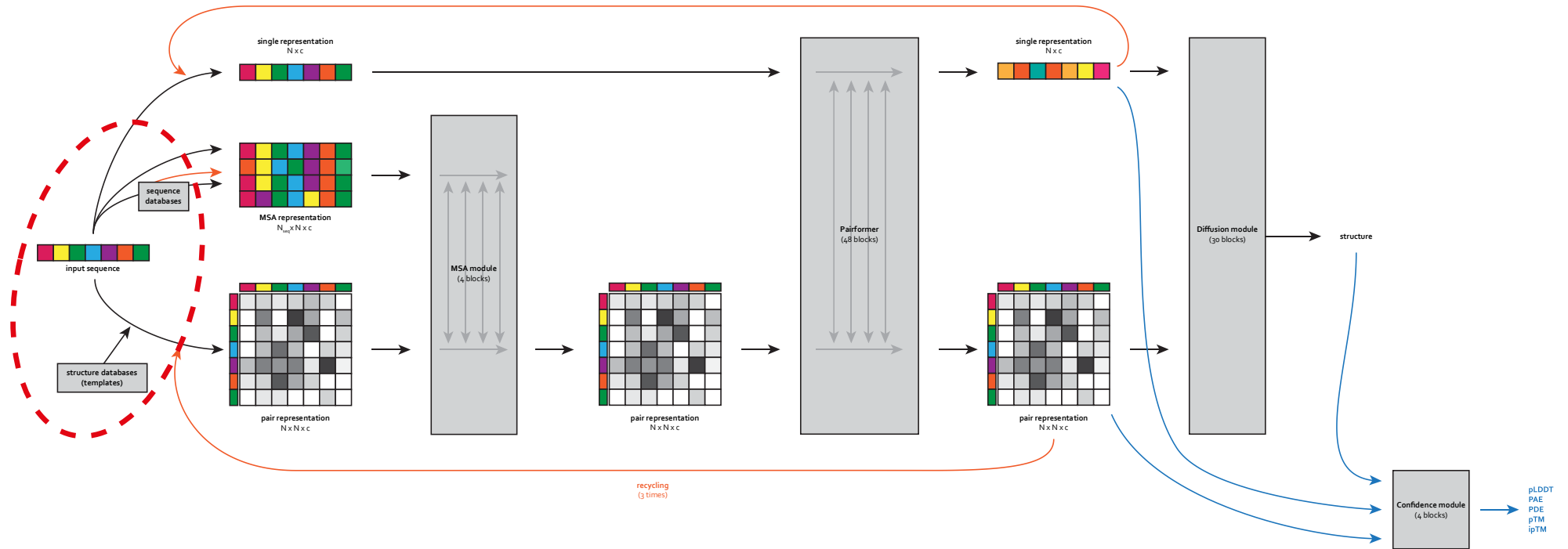


# AlphaFold2

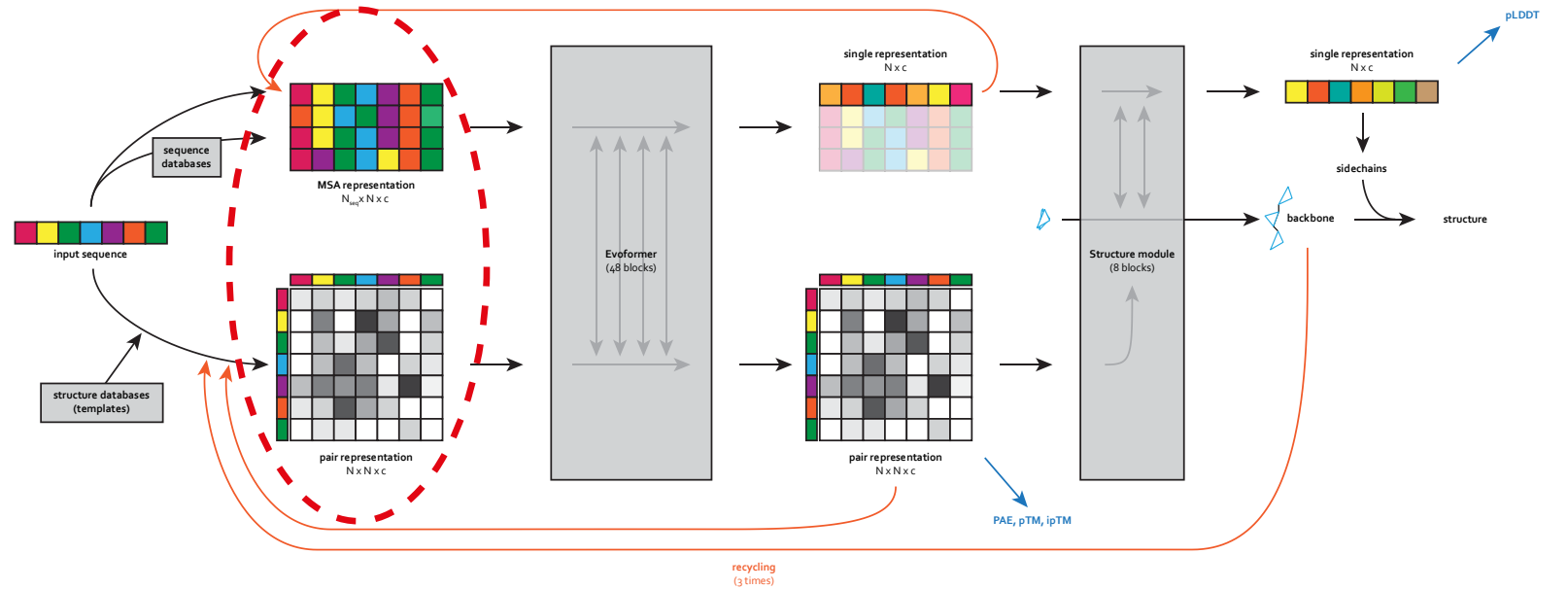


## Similar inputs

# AlphaFold3

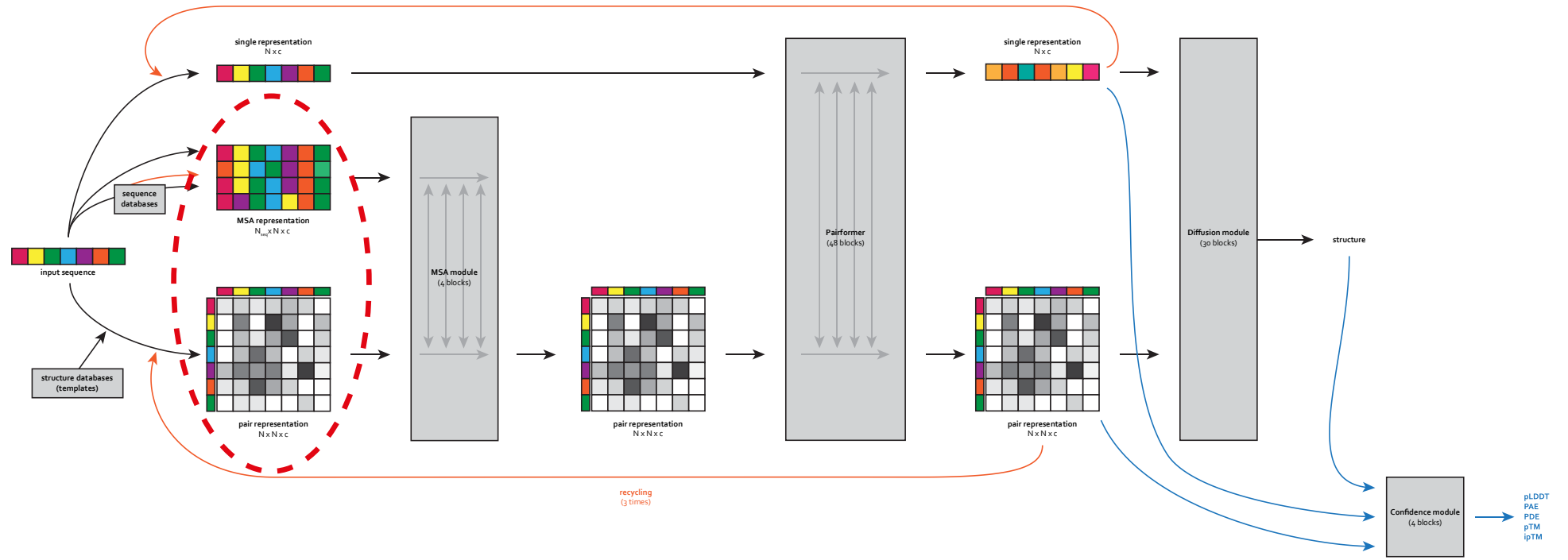


# AlphaFold2

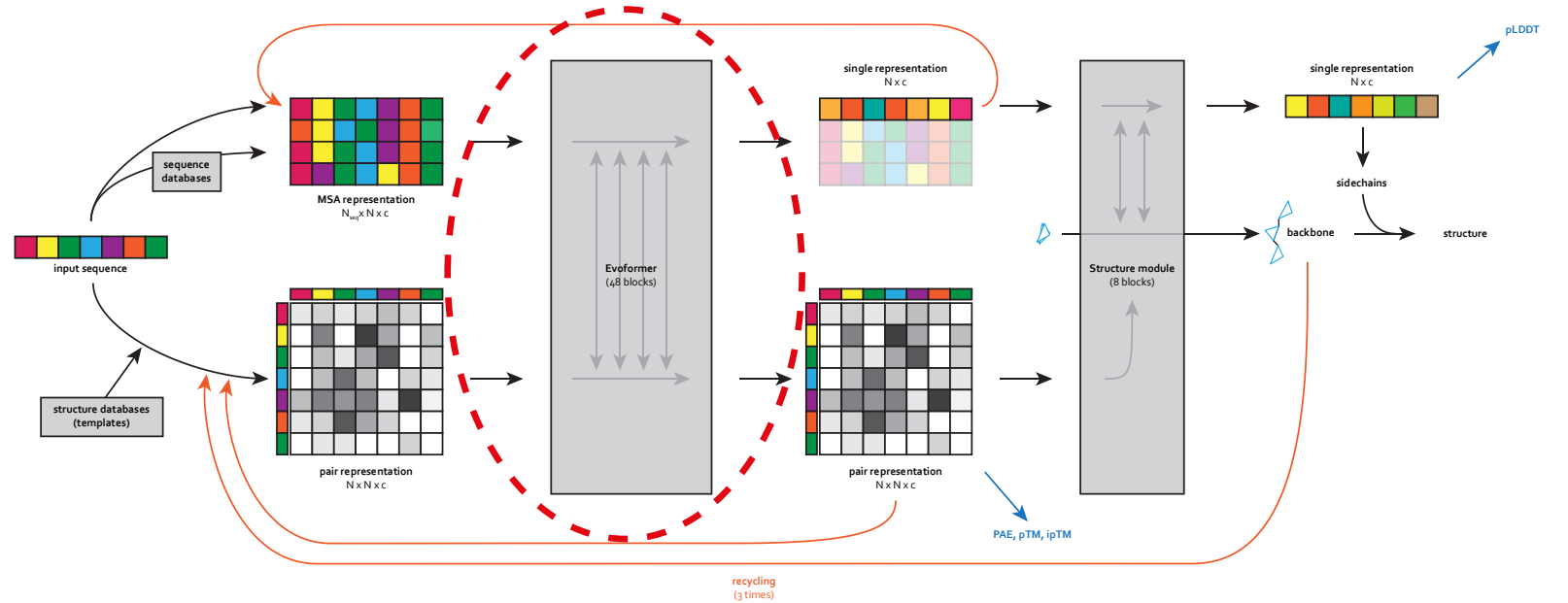


## Similar representations

# AlphaFold3

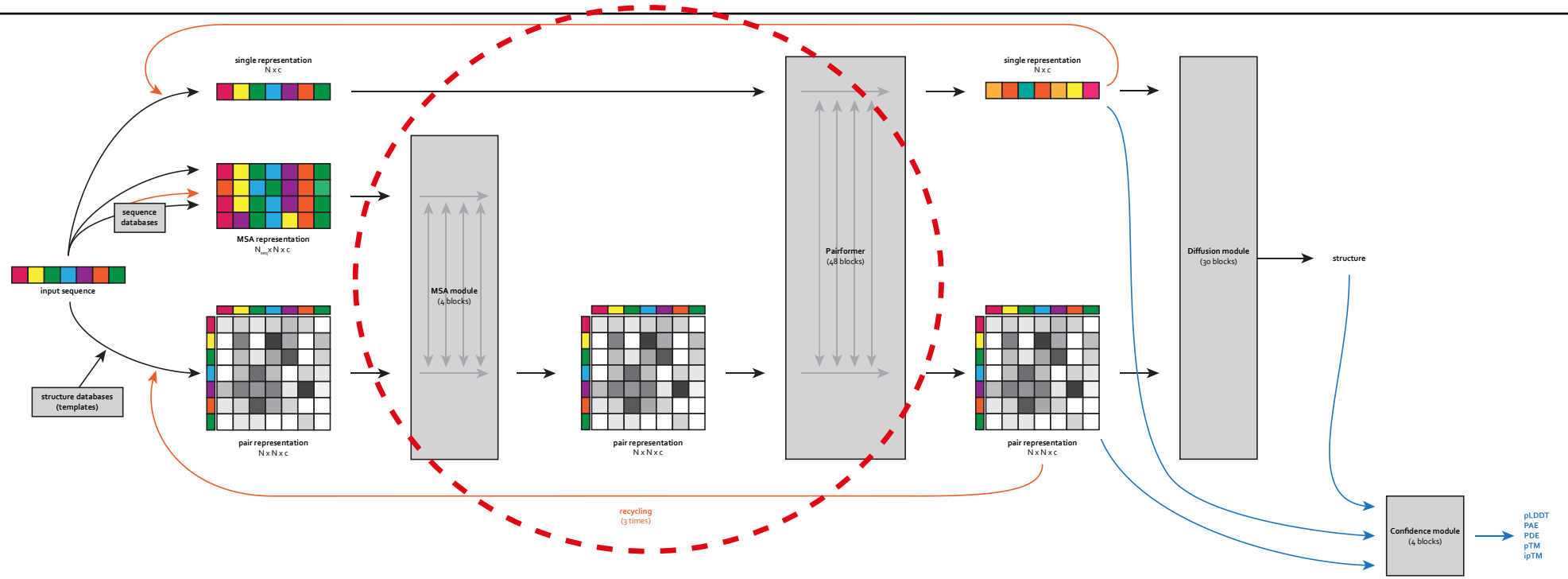


# AlphaFold2

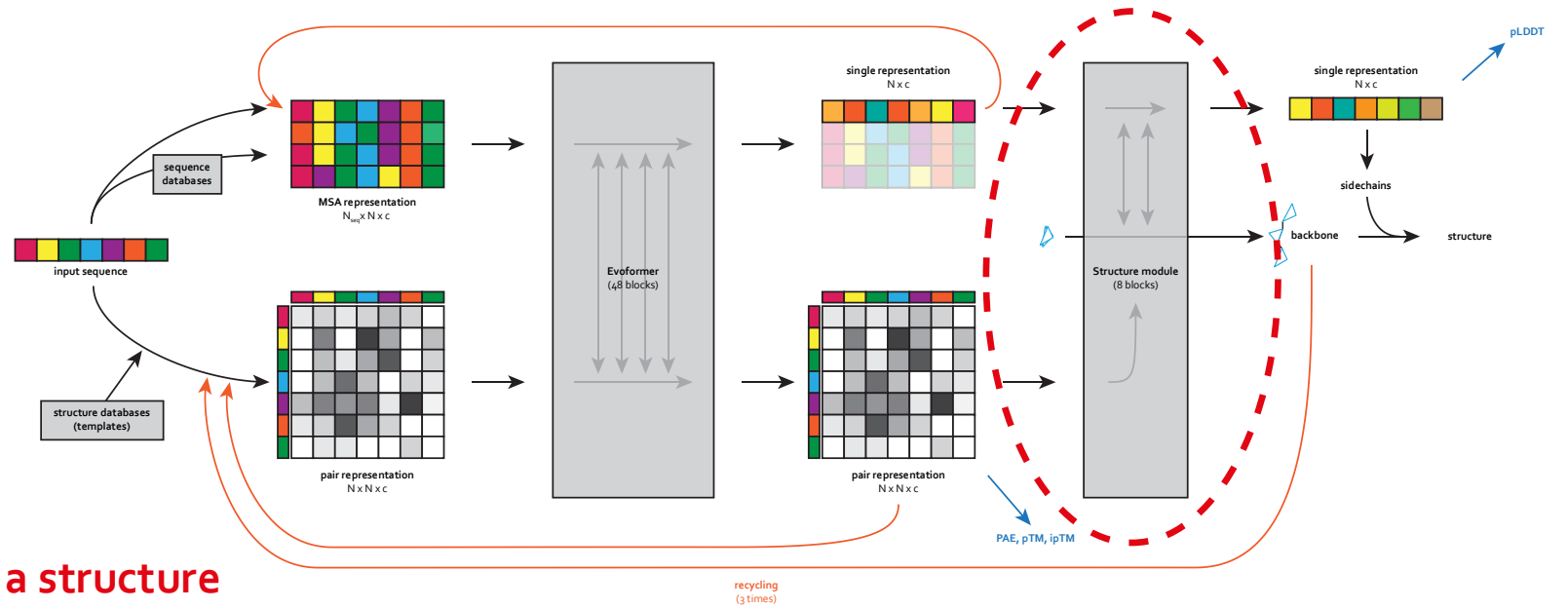


## Different MSA processing

# AlphaFold3

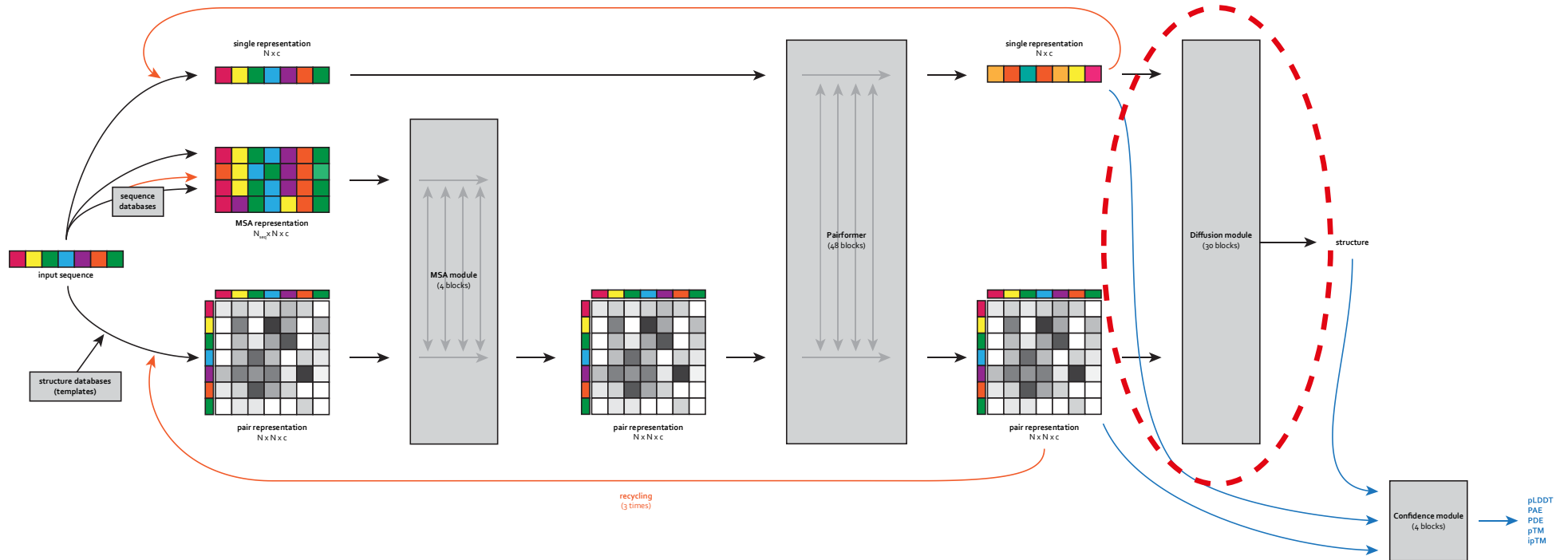


# AlphaFold2



Different way of generating a structure

# AlphaFold3





# Structure/Diffusion module

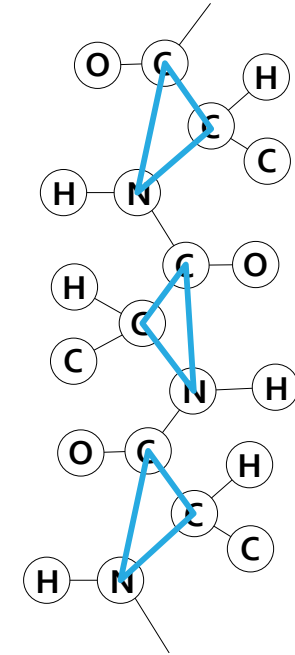
## AlphaFold2

(Structure module)

- Backbone frames
- Starting at 0
- Rotations, translations



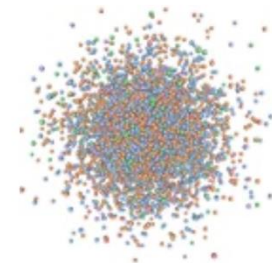
translations, rotations →



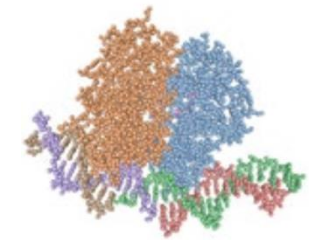
## AlphaFold3

(Diffusion module)

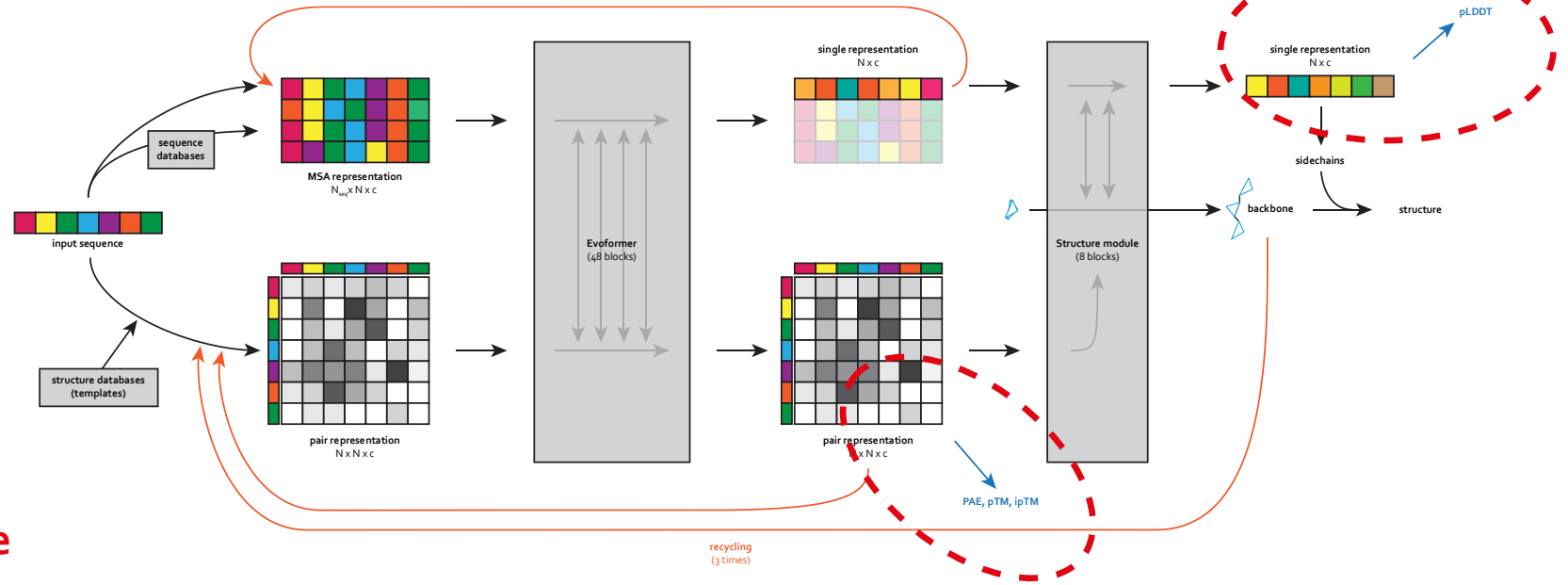
- Coordinates of all atoms
- Starting from random coordinates
- Denoising task



denoising →

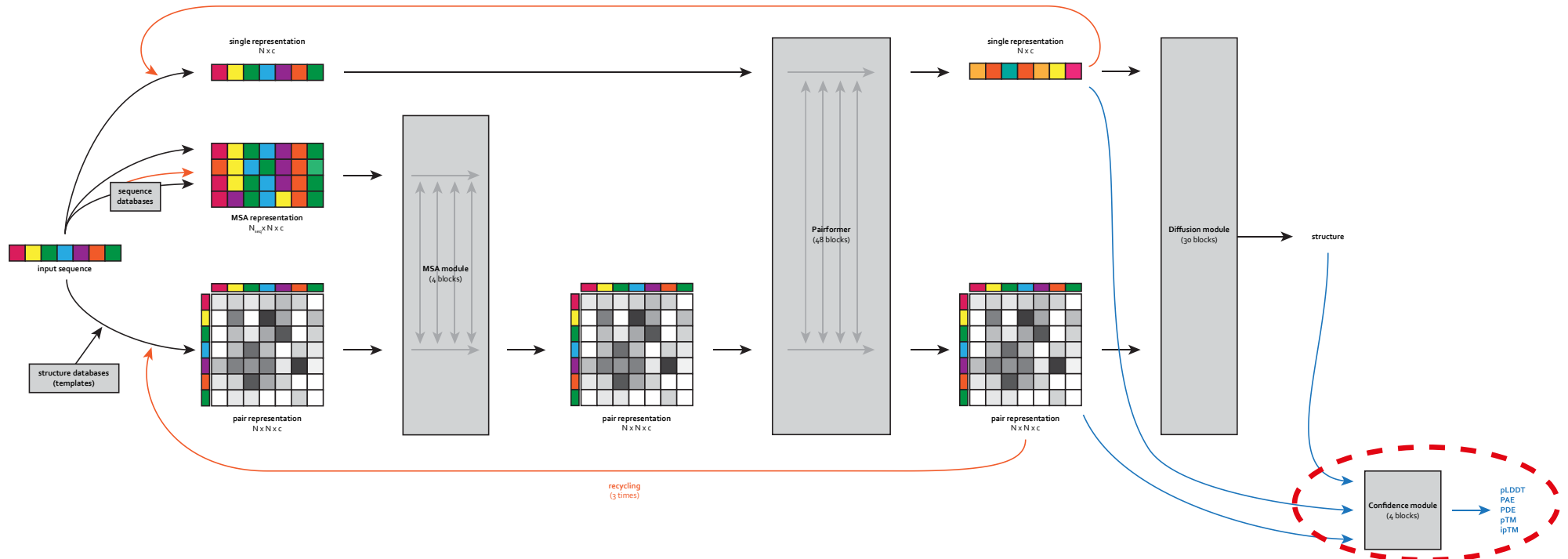


# AlphaFold2



## Different confidence module

# AlphaFold3

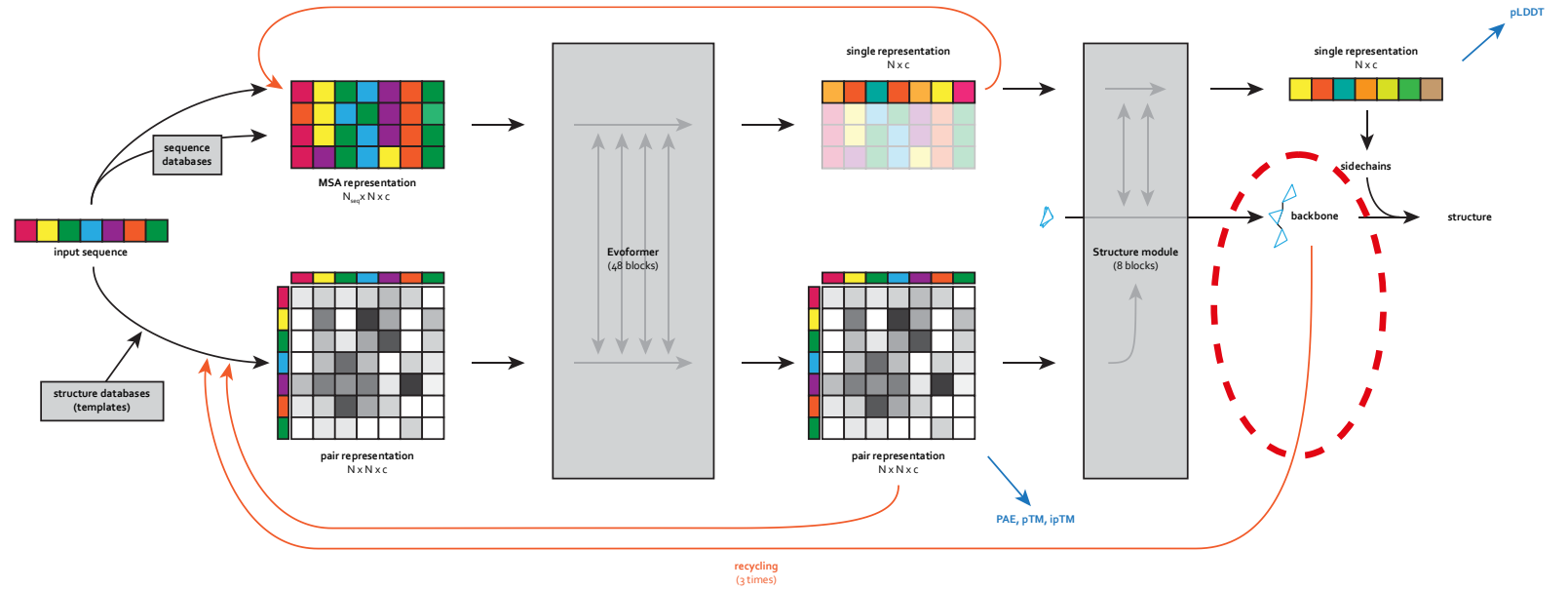




# Confidence scores

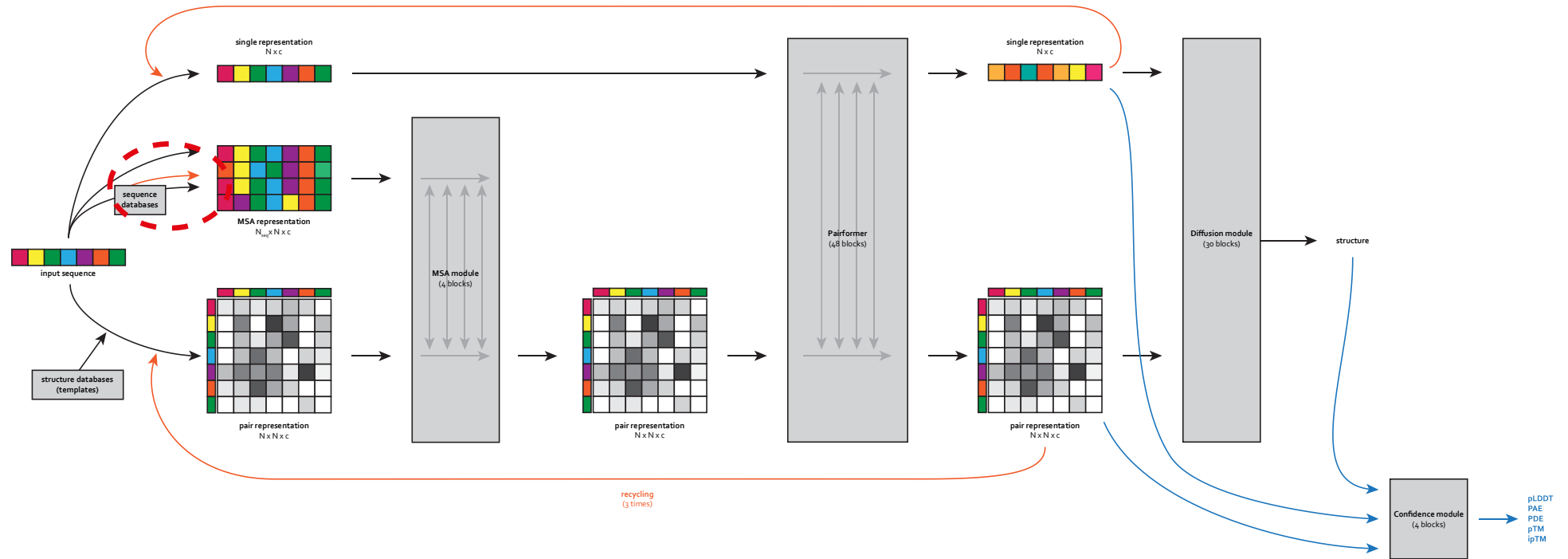
- The same metrics
- Calculated per-token
- PDE – predicted distance error
  - Predicted error in distance between two tokens (slightly different than PAE)
- Chain pair ipTM

# AlphaFold2



Slightly different recycling

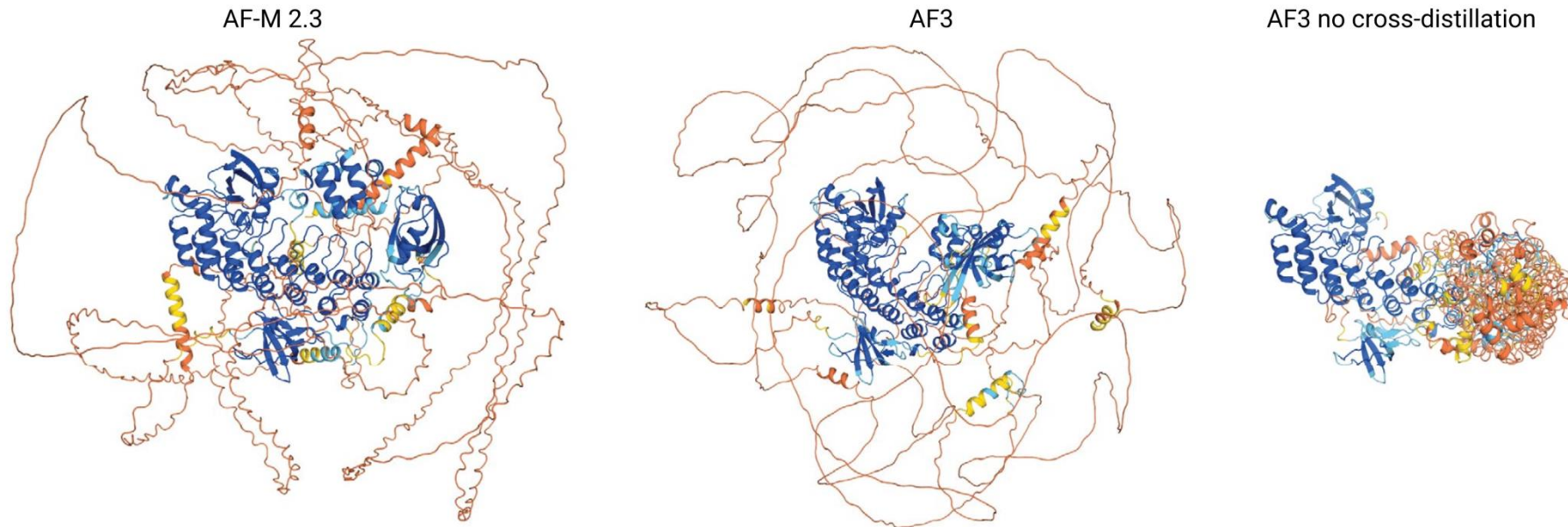
# AlphaFold3





# Hallucinations - structure in unstructured regions

- AlphaFold2 tends to represent these regions by long extended loops (low confidence + visual)
- AlphaFold3 might invent plausible-looking structure in these regions (still with low confidence, but people didn't like it visually)
- Cross-distillation: AF3 trained on AF2 structures to teach this behaviour

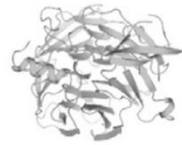




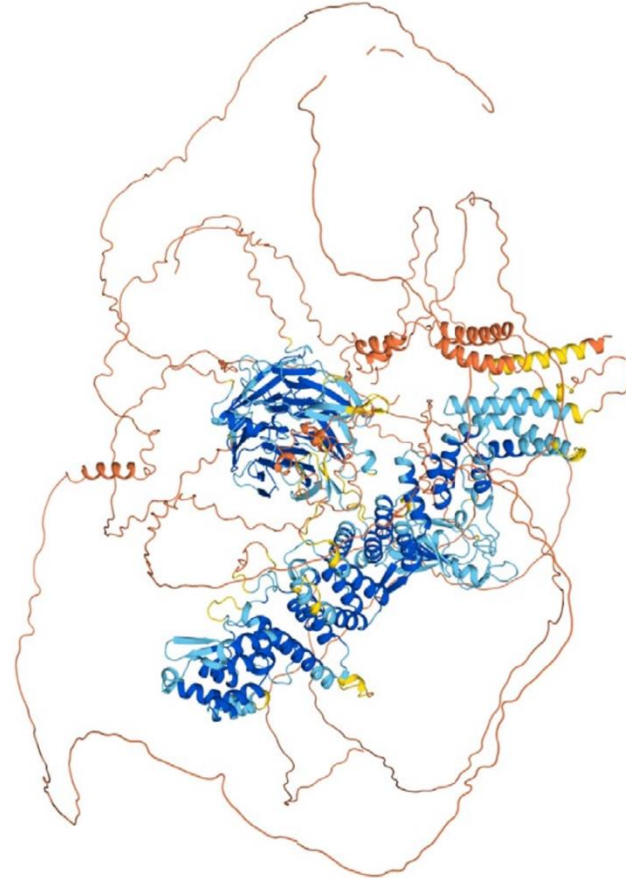
# Hallucinations - structure in unstructured regions

- Hallucinations can still appear

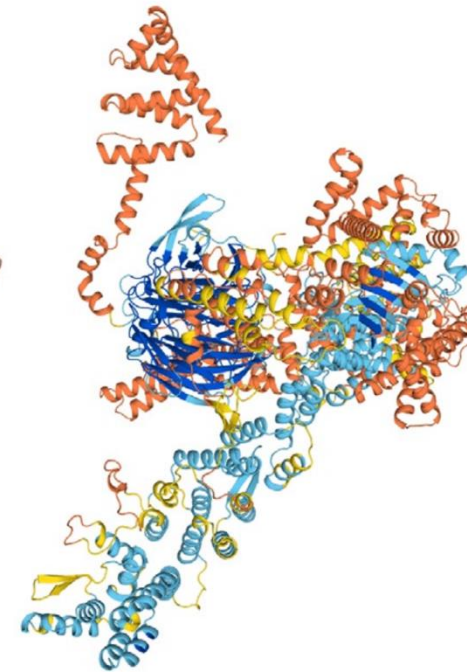
Ground truth



AF-M 2.3



AF3





# How good is it?



## **Proteins:**

similar to AlphaFold2



## **Protein-protein complexes:**

similar to AlphaFold2



## **DNA:**

not great, but the best for now



## **RNA:**

there are now better tools



# How good is it?

## Peptides:

work well

## Orphan proteins:

not great (similar as AlphaFold2)

## Antibodies:

work well, but require many samples (~500)

→ also check ESMFold2

## Viruses:

reasonable results for host–pathogen interactions (including viruses)<sup>[1]</sup>

[1] <https://pmc.ncbi.nlm.nih.gov/articles/PMC12258915/>

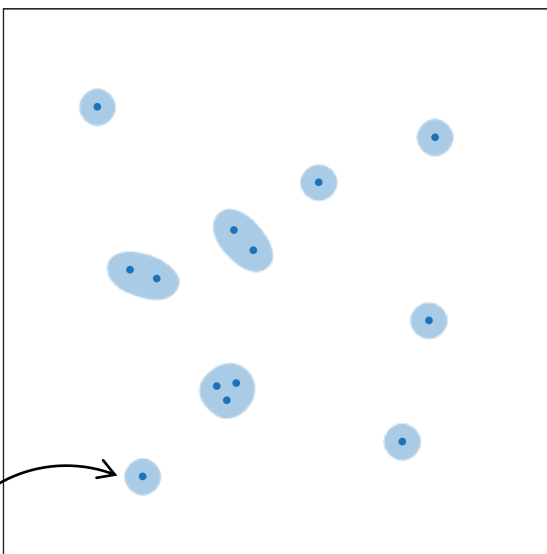
[2] [https://colab.research.google.com/github/Biohub/esm/blob/main/cookbook/tutorials/esmc\\_mutation\\_scoring.ipynb](https://colab.research.google.com/github/Biohub/esm/blob/main/cookbook/tutorials/esmc_mutation_scoring.ipynb)



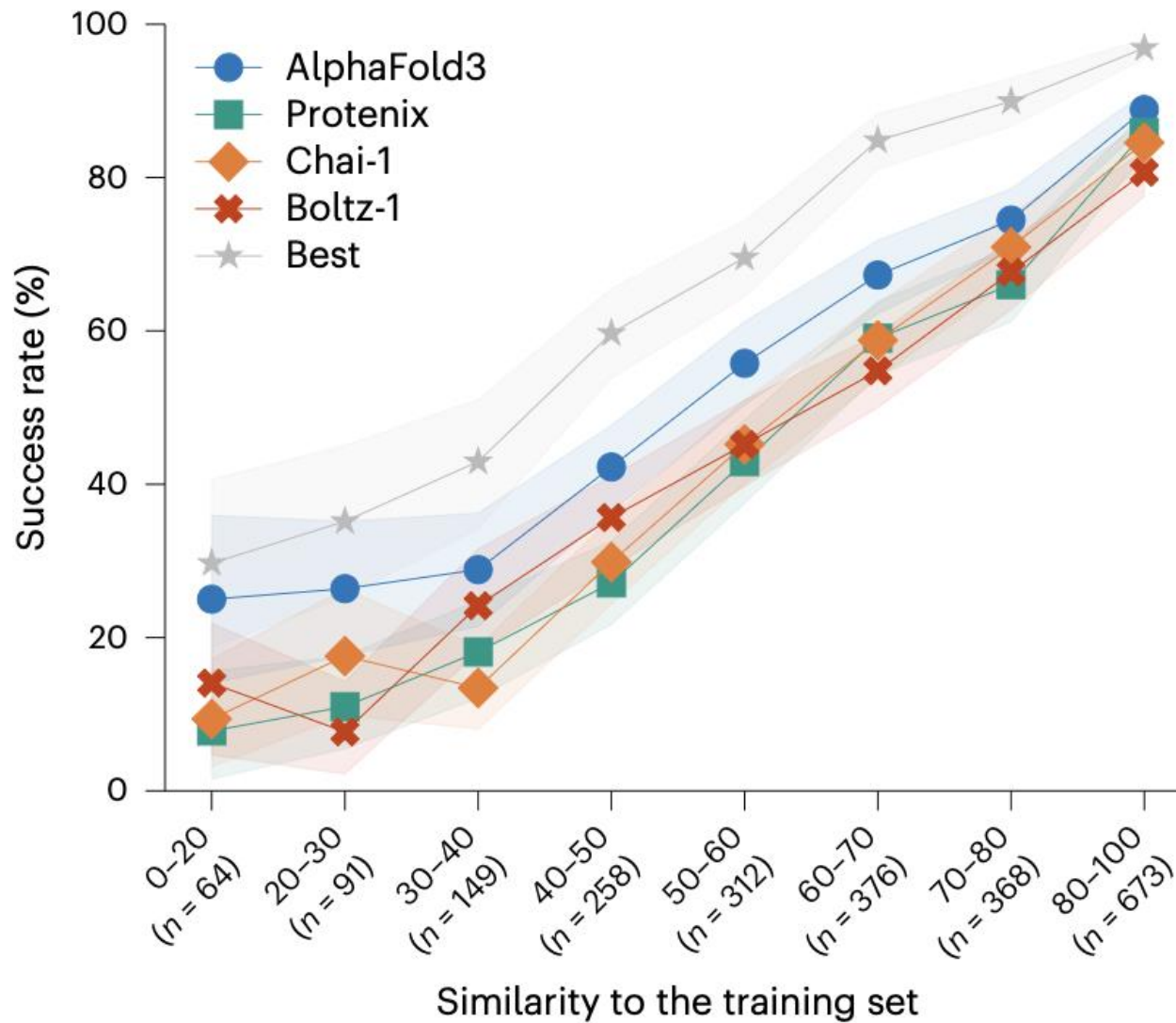
# Protein-ligand complexes



- Poor performance
- No such thing as MSA for protein-ligand interaction



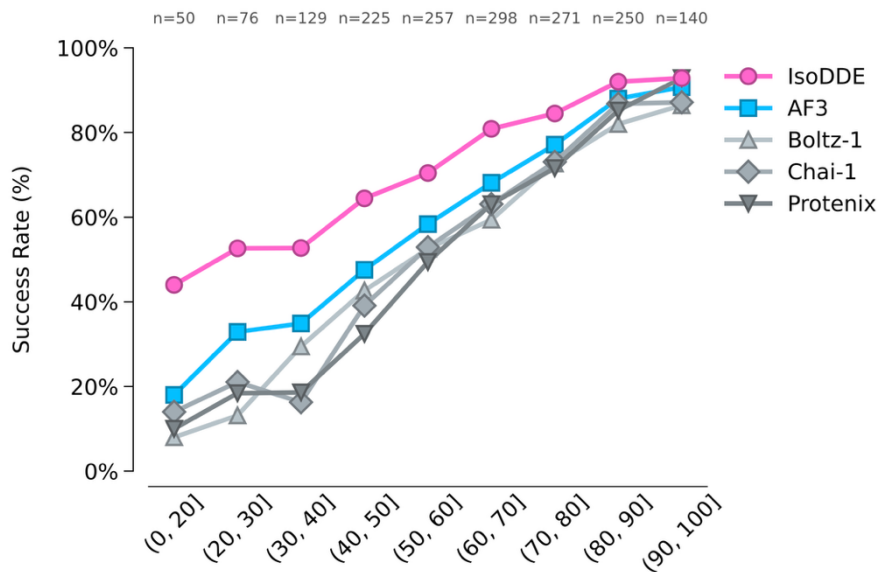
Known structures



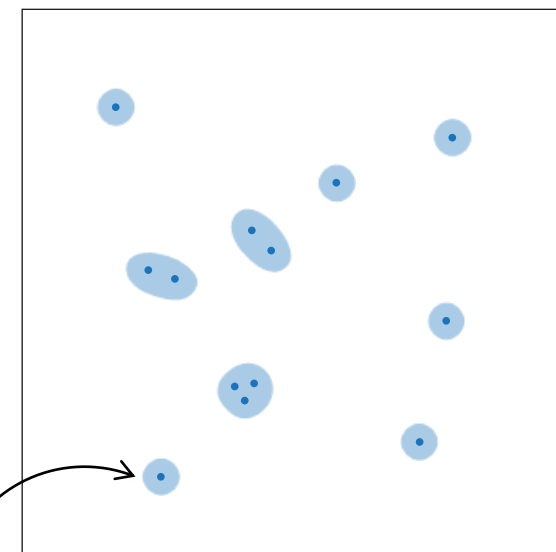


# Improving the performance on protein-ligand complexes

- More experimental structure data to train on<sup>[1]</sup>
  - Will take years
- Synthetic data<sup>[2]</sup>
  - Generated using physics-based methods



Pocket and ligand shape similarity to the training set



Known structures

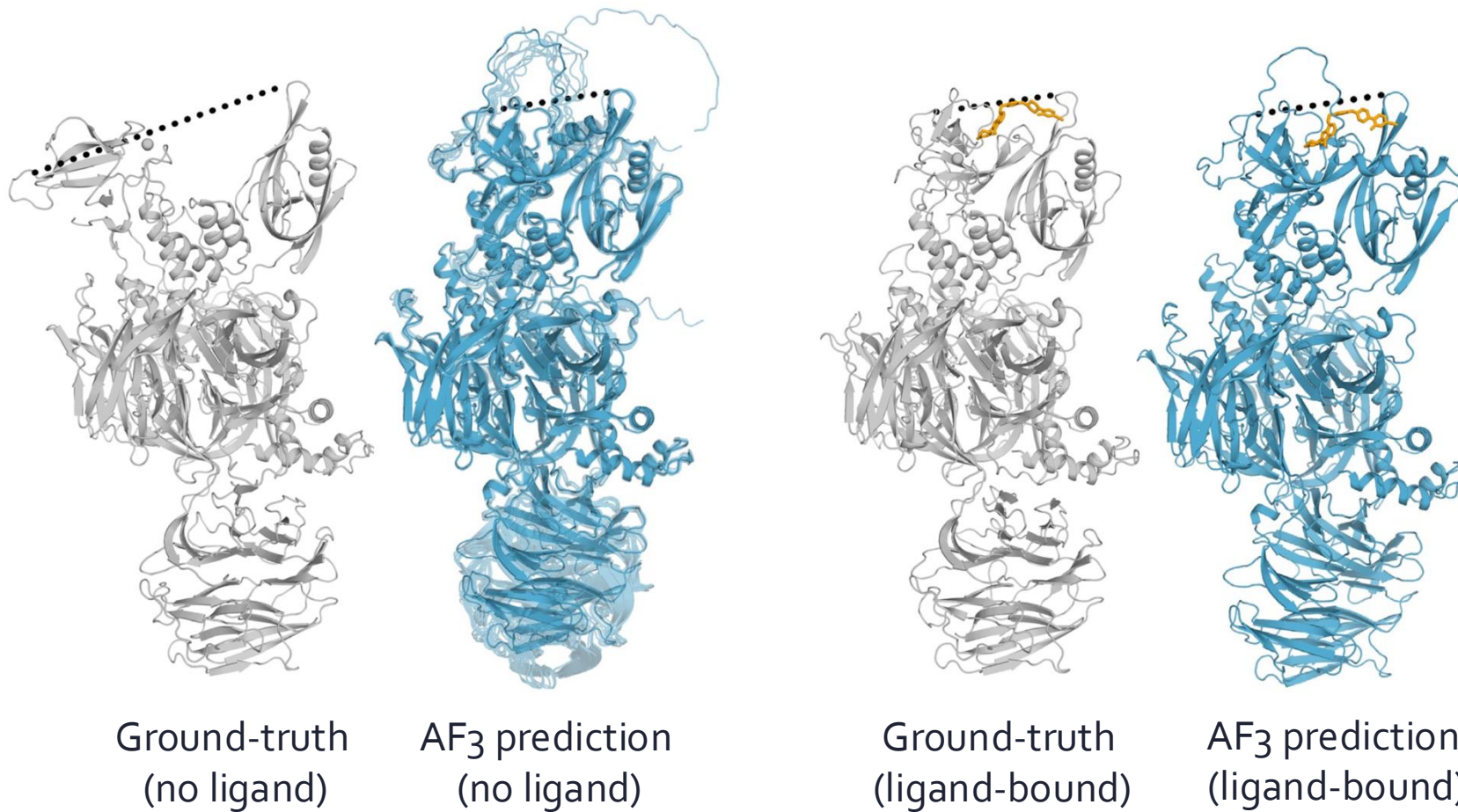
[1] <https://openbind.uk>

[2] <https://arxiv.org/abs/2510.24670v2>



# Limitations

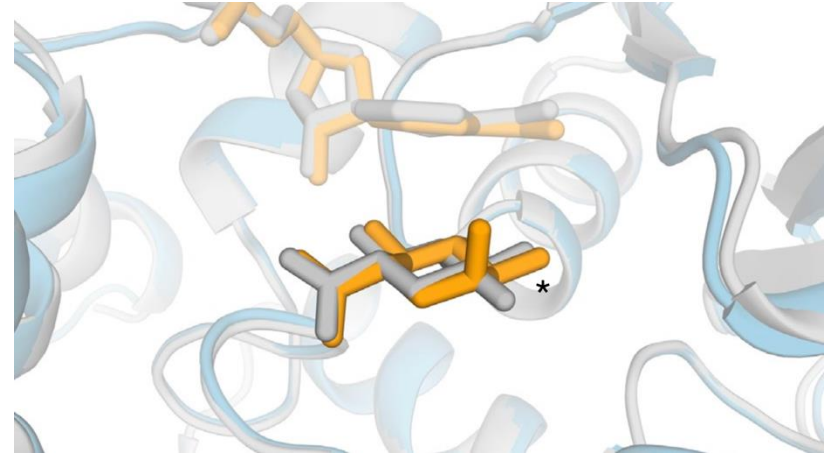
- Conformation coverage is limited





# Limitations

- Stereochemistry (chirality) violations
  - Not the one we specified as the input
- Confidence scores are overestimated
  - Users prefer AlphaFold2 metrics for some applications





# Which one should I use? (AF2 or AF3)

- AF2 is freely available for both academic and commercial use  
AF3 is available for non-commercial projects only
- Custom MSA/template mode is easier in AF3 compared to AF2 (for the local version)
- Online versions:  
ColabFold (AlphaFold 2) allows you to adjust parameters;  
the AlphaFold Server does not offer as much flexibility;
- AF2 is better for de novo proteins (due to better confidence metrics);
- Use AF3 if you want to model ligands, DNA, RNA, or Post-translational modification (PTM) – possible but not great
- AF3 is better for antibody-antigen complex prediction



# How to access

- Source code
  - Hard
  - Licence limitations
- AlphaFold Server
  - Online
  - No SMILES input
- A lot of “clones” exist, some include extra features:
  - Boltz (Boltz-1, Boltz-1x, Boltz-2, Boltz-2x)
  - Chai-1
  - ...



Swiss Institute of  
Bioinformatics

DAY 2, PART 3

# AlphaFold Server tutorial

Diana Rapota, Rok Breznikar, Janani Durairaj

23-24 June 2026





# AlphaFold Server

- Free and you do not need to install any software on your computer
- Supports modelling of multi-entity complexes (proteins, DNA, RNA, ligands, ions), but exposes only a limited subset of the full AlphaFold 3 (check FAQs)
- The minimum sequence length is four amino acids or four nucleotides
- Each modelling job is limited to 5,000 tokens
- Each user is limited to 30 jobs per day

AlphaFold Server [Server](#) [About](#) [FAQ & Guides](#) ▾

Remaining jobs: 30

AlphaFold Server allows you to model a structure consisting of many biological molecules

[Learn more](#) ^

- Remaining jobs refresh each day
- Jobs can be up to 5,000 tokens - see more details on token calculation, accepted formats, seed selection and other features in our [FAQ](#)
- Use the entity bar to chemically modify proteins and nucleic acids
- Get in touch with the AlphaFold team if you have any questions

Explore these examples of structures to see it in action – try them out without using your quota until you begin editing!

Protein-RNA-Ion: PDB 8AW3

Protein-Glycan-Ion: PDB 7BBV

Protein-DNA-Ion: PDB 7RCE

Ok, got it



# AlphaFold Server: Settings

- There are options for custom templates and MSA
- You can specify a cut-off date for the PDB templates to be used. The cut-off date for the template is usually used if you don't want templates that AF3 was trained on (training cut-off date: 2021-09-30).

## Template settings

Use PDB templates up to

Use PDB templates with default cut-off date (30/09/2021)

Turn off templates

Cancel

Save

## Custom MSA

Use internally computed MSA

Cancel

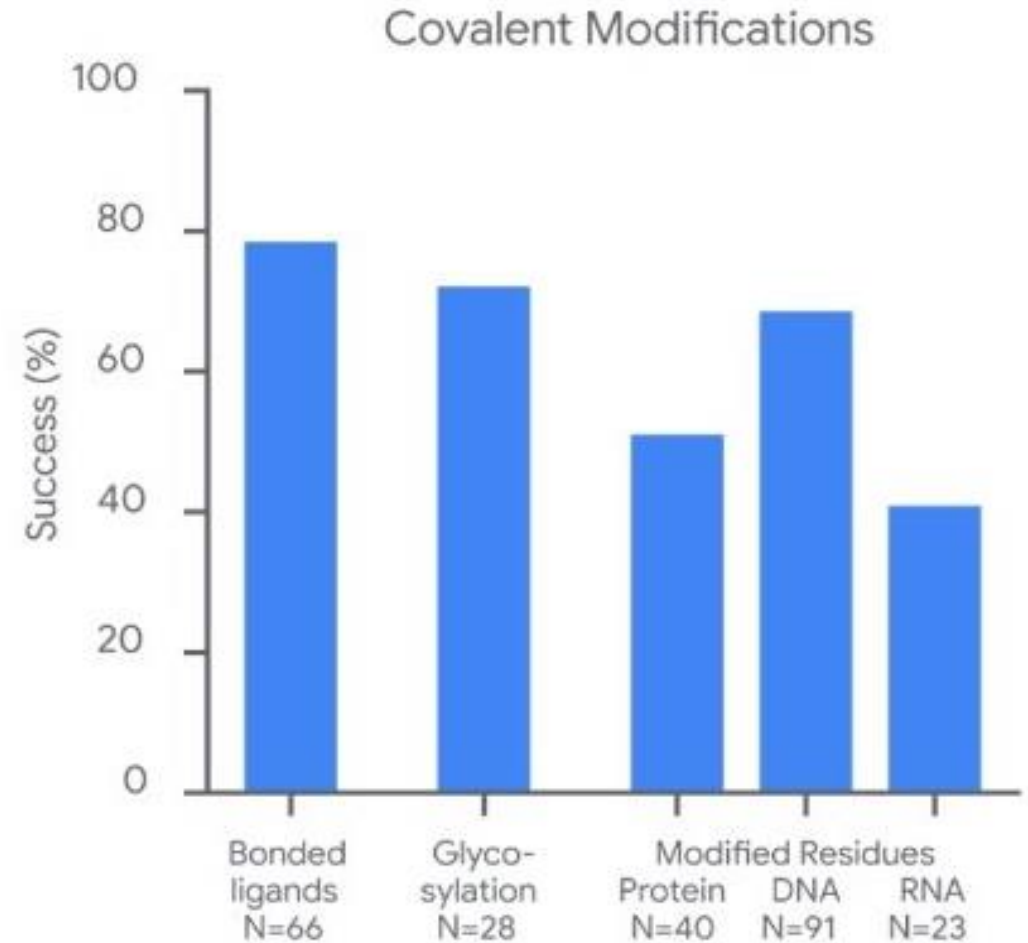
Save



# AlphaFold Server: Settings

## Adding modifications

- Multiple modifications to different residues one at a time (for Glycosylated proteins see AlphaFold Server FAQ)
- Modifications for DNA and RNA



*Abramson et al, 2024*



# AlphaFold Server: Advanced Settings

- **JSON job submission** for automatic job generation (useful for screening experiments, up to 100 jobs per file; you are allowed up to 500 saved drafts)
- **Sampling multiple seeds** (20 seeds is usually enough to get a better prediction)
- **Reproducing jobs** (just use the same input and seed number, which can be found in `fold_<job_name>_job_request.json` - contains all the information required for reproduction)



# AlphaFold Server: Output

- Five .cif files named **fold\_<job\_name>\_model\_<N>.cif** contain predicted structures in the mmCIF format (can be viewed in any molecular viewer);
- Five .json files named **fold\_<job\_name>\_summary\_confidences\_<N>.json** contain summaries of the confidence metrics for the predictions;
- Five .json files named **fold\_<job\_name>\_full\_data\_<N>.json** contain detailed confidence metrics (per-atom pLDDT, per-token contact prob matrix, per-token pae etc.)
- A file named **fold\_<job\_name>\_job\_request.json** contains the inputs of the modelling job and could be used to re-run the job;
- A file named **terms\_of\_use.md**. This is a legal document detailing the terms of use for the predictions.

where N is the rank of the predicted structure, structures are ranked from 0 to 4, where 0 has the highest confidence



# AlphaFold Server: Confidence metrics

JSON files with **summary** contain:

- **chain\_ipTM**: the average confidence (ipTM) in the interfaces between each chain and all other chains, can be used for ranking predicted structures for a specific chain;
- **chain\_pair\_ipTM**: pairwise ipTM scores, can be used for ranking predictions of a structure by the accuracy of a specific interface between two chains;
- **chain\_pair\_pae\_min**: element (i, j) of the array contains the lowest PAE value across rows restricted to chain i and columns restricted to chain j. This has been found to correlate with whether or not two chains interact;
- **chain\_ptm**: pTM restricted to chain i;
- **fraction\_disordered**: range 0-1 that indicates what fraction of the prediction structure is disordered;
- **has\_clash**: 1.0 if more than 50% of a chain's atoms clash, or if a chain has more than 100 clashing atoms; 0.0 otherwise.
- **iptm**: ipTM for all interfaces in the structure;
- **ptm**: pTM-score for the full structure;
- **ranking\_score**:  $0.8 \times \text{ipTM} + 0.2 \times \text{pTM} + 0.5 \times \text{disorder} - 100 \times \text{has\_clash}$ , range from -100 to 1.5



# AlphaFold Server: Confidence metrics

JSON files with **full outputs** contain:

- **atom\_chain\_ids**: the chain IDs corresponding to each atom in the prediction;
- **atom\_plddts**: A [num\_atoms] array, element *i* indicates the pLDDT for **atom** *i* in the prediction;
- **contact\_probs**: A square [num\_tokens, num\_tokens] array. Element (*i*, *j*) indicates the predicted probability that **token** *i* and **token** *j* are in contact;
- **paе**: A square [num\_tokens, num\_tokens] array. Element (*i*, *j*) indicates the PAE in the position of **token** *j*, when the prediction is aligned to the ground truth using the frame of **token** *i*;
- **token\_chain\_ids**: A [num\_tokens] array indicating the chain IDs corresponding to each token in the prediction;
- **token\_res\_ids**: A [num\_res] array.